Methanol / Chloroform Extraction for Proteins

Guidelines for sample preparation
(How to protect your samples from contamination with keratin)

- TRY TO AVOID ANY CONTACT OF SAMPLES AND SOLUTIONS WITH DUST, SKIN OR HAIR
- CLEAN YOUR BENCH
- WEAR GLOVES AT ALL TIMES
- ALL REAGENTS SHOULD BE PREPARED FRESH
- USE ULTRA PURE WATER FOR ALL SOLUTIONS

SAMPLE SUBMISSION

- PROVIDE SAMPLES IN 1.5 ML TUBES (NO SCREW CAPS)
- LABEL YOUR TUBE WITH YOUR NAME, SAMPLE NUMBER AND DATE
- PLEASE FILL IN ONLINE SAMPLE SUBMISSION FORM TO PROVIDE US WITH MORE INFORMATION ABOUT YOUR SAMPLE (PROTEIN ORIGIN, TAXONOMY, CONCENTRATION)

- Use 200 µl of sample. If sample volume is bigger than 200 µl, split into multiple tubes. If sample is less than 200 µl bring the volume up to 200 µl with MilliQ-H₂O.
- Add 600 µl methanol
- Add 150 µl chloroform
- VORTEX
- Add 450 µl MilliQ-H₂O
- VORTEX
- Centrifuge at room temp. for 1 min.
- Pipette off upper aqueous phase without disrupting the precipitate at the interface.
- Add 450 µl methanol
- VORTEX
- Centrifuge at room temp. for 2 min.
- Remove supernatant
- go to “IN-SOLUTION DIGESTION”